

REVIEW ARTICLE



Controversies in Pathology

Breslow thickness 2.0: Why gene expression profiling is a step toward better patient selection for sentinel lymph node biopsies

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Risk-stratification of cutaneous melanoma is important. Patients want to know what to expect after diagnosis, and physicians need to decide on a treatment plan. Historically, melanoma that had spread beyond the skin and regional lymph nodes was largely incurable, and the only approach to preventing a bad outcome was surgery. Through the seminal work of Alexander Breslow and Donald Morton, a system was devised to carefully escalate surgery based on primary tumor thickness and sentinel lymph node status. Today, we know that prophylactic lymph node dissections do not improve survival, but we continue to appreciate the prognostic implications of a positive sentinel node and the benefits of removing nodal metastases, which facilitates locoregional disease control. However, the question arises whether we can better select patients for sentinel lymph node biopsies (SLNB) as, currently, 85% of these procedures are negative and non-therapeutic. Here, we argue that gene expression profiling (GEP) of the diagnostic biopsy is a valuable step toward better patient selection when combined with reliable clinicopathologic (CP) information such as patient age and Breslow thickness. Recently, a CP-GEP-based classifier of nodal metastasis risk, the Merlin Assay, has become commercially available. While CP-GEP is still being validated in prospective studies, preliminary data suggest that it is an independent predictor of nodal metastasis, outperforming clinicopathological variables. The hunt is on for Breslow thickness 2.0.

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BACKGROUND

Sentinel lymph node biopsy (SLNB) in cutaneous melanoma is critical for therapeutic decision making^{1,2}. Initially, when the method was devised, a positive SLNB signaled the need for a completion lymph node dissection (CLND)³. At the time, effective systemic adjuvant therapy was unavailable, and lymphadenectomy was the dominant of a handful of feasible interventions. Surgical oncologists, who saw their patients relapse and die from unstoppable disease, sought to intervene early and aggressively, following the adage that a chance to cure is a chance to cure⁴. Eventually, both the Multicenter Selective Lymphadenectomy Trial (MSLT)-2 and Dermatological Cooperative Oncology Group (DeCOG)-SLT trial failed to confirm a survival benefit for CLND in SLNB-positive patients^{5,6}. While CLND may still have a role in selected patients⁷, the procedure is no longer standard treatment.

Today, SLNB is used to rule in adjuvant therapies¹. Still, not all patients are believed to be high enough risk for the procedure. Per current guidelines, SLNB is not recommended if the risk of nodal metastasis is <5%, as in T1a melanoma with a Breslow thickness of <0.8 mm and no adverse features. SLNB should be considered if the risk of nodal metastasis is between 5% and 10% (T1b, Breslow thickness, 0.8–1.0 mm) and is recommended if the risk of nodal

metastasis exceeds 10% (T2 to T3, Breslow thickness, >1.0 mm)^{1,2}. Patients with T3b and T4 melanoma (Breslow thickness of >2.0–4 mm with ulceration or Breslow thickness >4 mm) qualify for adjuvant therapy irrespective of nodal status and may forgo the procedure⁸. Of all SLNB-eligible patients, less than 20% have nodal metastasis^{9,10}. When performed by a skilled surgeon, the SLNB procedure is safe, but patients face a risk of surgical complications, including bleeding, infection, seroma, and lymphedema^{11,12}.

While SLNB is an important tool for individualizing therapy, the method has several disadvantages. The false-negative rate has been estimated at 15–20%^{9,13–15}, which is consistent with our experience, and is due to technical problems, operator-dependency, disruption of lymphatic drainage from diagnostic biopsies, idiosyncratic lymphatic obstruction, complex lymphatic drainage in the head and neck, inadequate histological analysis and complex metastasis patterns. SLNB can also be false-positive when benign melanocyte-marker-positive cells in sentinel nodes are called malignant¹⁶. Up to 5.1% of lymph nodes from patients with no history of melanoma contain single MART-1 positive cells¹⁷. In a series of T1 to T3 cutaneous melanoma from Mayo Clinic, 24.3% of positive SLN showed only individual melanocytes or melanocytic cell clusters <0.1 mm¹⁰. Differentiating single benign from malignant melanocytes is challenging, especially

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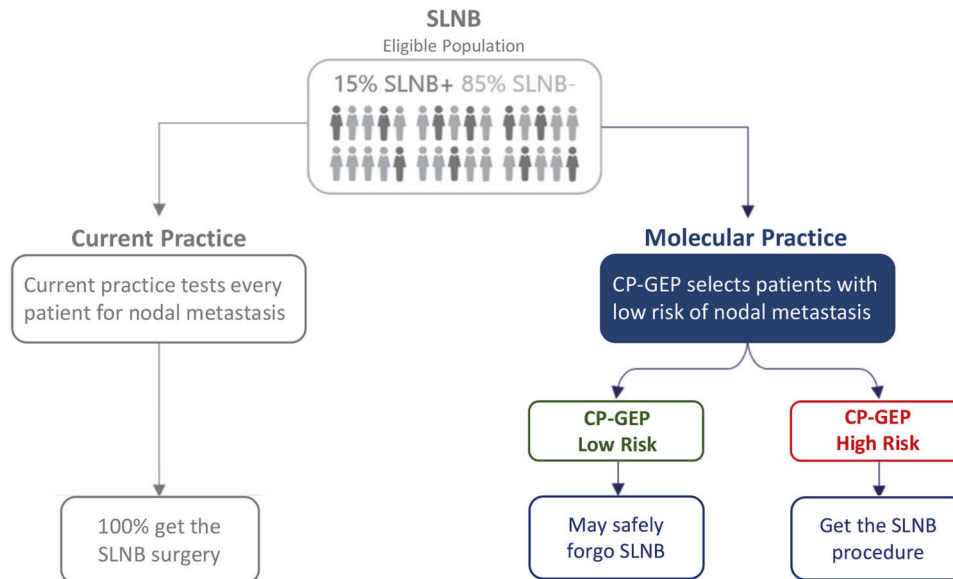


Fig. 1 Based on current patient selection, approximately 85% of sentinel lymph node biopsies (SLNB) are negative (SLNB-) and non-therapeutic. In the future, molecular tests based on gene expression profiling (GEP) may be combined with clinicopathologic (CP) variables such as Breslow thickness and patient age to better select patients for SLNB. CP-GEP low risk patients can forgo SLNB without compromising oncologic safety.

when cells are only found on immunohistochemistry sections but not on corresponding hematoxylin eosin sections¹⁶.

When Donald Morton devised the SLNB procedure for cutaneous melanoma in the 1990s, molecular diagnostics was in its infancy. Routine molecular analysis of paraffin-embedded diagnostic biopsy tissue was not feasible. The paradigm shifted after 2000, when OncotypeDx¹⁸ and MammaPrint¹⁹ were introduced, gene expression-based assays that stratified the likelihood of breast cancer recurrence. Today, we can apply a vast machinery of sophisticated molecular tools to analyze routinely processed biopsy samples. The question arises whether a complex surgical procedure like SLNB²⁰ can be supplemented by the molecular analysis of diagnostic biopsy tissue, for example, to better select patients for the procedure (Fig. 1).

CLINICOPATHOLOGIC PREDICTORS OF METASTASIS RISK

Breslow thickness is the bedrock on which melanoma staging is built². Conceived in the late 1960s by Alexander Breslow²¹, the idea of using tumor thickness as a predictor of metastasis and survival is powerful and simple. Measuring Breslow thickness accurately, however, requires the complete excision of the primary tumor, which in today's busy clinical practice does not always take priority. There is an increase in the use of shave biopsies, which are fast, easy to do, and sutureless, but can lead to transected tumors^{22,23}. Other variables have stood the test of time. Paradoxically, numerous studies have found that younger age increases both SLNB metastasis risk and survival rates²⁴, whereas elderly patients are less likely to be node positive but have a worse prognosis²⁵. In pediatric patients, SLNB loses its prognostic value because sentinel nodes are frequently positive and the clinical course is excellent²⁶.

Additional primary tumor-derived prognostic variables include tumor ulceration, Clark level, mitotic rate (MR), lymphovascular invasion (LVI), tumor-infiltrating lymphocytes (TILs), and histologic type, among others. While these have been known for decades and studied extensively, they have limitations. For example, the consistency of reporting Clark level, MR, LVI, and TILs is low, highlighting problems with reliability²⁷. Discordance in the reporting of MR between institutions is high and can result from

the counting of mitotic figures at the dermo-epidermal junction instead of the invasive component of a melanoma or by counting mitotic non-melanocytes²⁸. Problems with reliability have led to the removal of MR as a staging variable for thin melanomas, a decision widely applauded and cited as the reason for the greater reliability of current guidelines²⁹. Other histologic features have powerful implications but are rare. Pure desmoplastic melanomas, for example, almost never metastasize to sentinel nodes but occur in only 3% of SLNB-eligible patients¹⁰. Acral melanomas often metastasize to sentinel nodes but occur in only 1–2% of SLNB-eligible patients in the United States^{10,30}.

Which clinicopathologic (CP) variables are used to make predictions? The American Joint Committee on Cancer (AJCC) relies on two primary tumor-derived histologic features for staging, Breslow thickness and tumor ulceration². The Memorial Sloan Kettering Cancer Center (MSKCC) nomogram for predicting SLNB outcome uses five CP variables: Breslow thickness, tumor ulceration, patient age, Clark level, biopsy location³¹. The more recent Melanoma Institute of Australia (MIA) nomogram considers six: Breslow thickness, tumor ulceration, patient age, histologic subtype, MR, and LVI³². The most ambitious models for predicting SLNB status, such as those developed by the University of Padova and MIA, combine up to eight CP variables, and put special emphasis on MR as a feature outperforming Breslow thickness³³.

A review of patients seen at Mayo Clinic did not reproduce the outsized role of MR as a predictor of nodal metastasis. When we applied statistical methods to identify CP-based predictors of nodal metastasis that maximize the separability of patients and minimize prediction error, Breslow thickness and patient age were sufficient¹⁰. More complex CP models did not improve model performance. We concluded that there is a limit to the ability of CP factors to predict SLNB status¹⁰. We further hypothesized that our vast new knowledge of cancer cell biology combined with the availability of new molecular techniques should facilitate the discovery of predictive and prognostic molecular features³⁴.

MOLECULAR PREDICTORS OF METASTASIS RISK

Ambitious CP models like those by Mocellin and coworkers³³ identified 18–30% of SLNB-eligible patients who could safely forgo

the procedure, meaning that the negative predictive value (NPV) of a low risk test result was greater than 90%. CP models, operationalized as nomograms, are therefore believed to hold promise as clinically relevant tools for SLNB decision making^{31,32}. To improve on the performance of CP variables, gene expression-based molecular biomarkers for melanoma risk-stratification have been explored in translational research^{34–37}, and are now introduced to clinical care^{10,38,39}. Current clinical guidelines acknowledge the potential of molecular tools, but also highlight the need for extensive prospective validation¹. Recently, a consensus statement by the Melanoma Prevention Working Group, a group of melanoma key opinion leaders, defined the required evidence to endorse the use of expression-based melanoma risk-stratification assays in clinical care⁴⁰. The group noted a lack of high-quality evidence supporting the routine use of prognostic gene expression profile (GEP)-based testing and highlighted the need for both prospective validation and benchmarking against CP variables.

Two prognostic GEPs are used in patient care or clinical trials, i.e., the 31-GEP (DecisionDx)³⁸ and 11-GEP (MelaGenix)⁴¹. 31-GEP is reimbursement by Medicare for identifying patients at risk of nodal metastasis⁴², but has been developed as a purely prognostic tool³⁸. 11-GEP is studied as a decision tool for postoperative adjuvant therapy in an ongoing multicenter trial for stage II melanoma in Germany⁴³. The added value of these GEPs over established CP variables has not yet been convincingly shown.

THE CP-GEP (MERLIN) TEST FOR MELANOMA RISK ASSESSMENT

Another concept of molecular testing was introduced with CP-GEP (Merlin)^{10,44,45}. The CP-GEP Assay, which is reimbursed by Medicare, is the only GEP-based classifier specifically developed to identify patients at risk of nodal metastasis. Unlike pure GEP-based models like 31-GEP or 11-GEP, the CP-GEP approach, introduced by Suman and colleagues³⁴, was to first develop models of the likelihood of nodal metastasis based on either CP variables (CP models) or GEPs of the primary tumor (GEP models) and then to assess the performance of a combined model of CP and GEP factors (CP-GEP models). CP variables considered included Breslow thickness, Clark level, MR, ulceration, LVI, biopsy location, regression, histologic type, and patient age at diagnosis. Of these, variable selection and regularization techniques selected Breslow thickness and patient age. More complex CP models did not improve performance. The combined CP-GEP model was based on Breslow thickness, patient age and the expression of eight well-characterized genes, which have been functionally linked to melanoma invasion and metastasis, i.e., Melanoma Antigen Recognized By T-Cells 1 (*MLANA*), Macrophage Inhibitory Cytokine 1 (*GDF15*), Interleukin 8 (*CXCL8*), Lysyl Oxidase Like 4 (*LOXL4*), TGF- β Receptor 1 (*TGFBR1*), β 3 Integrin (*ITGB3*), Tissue-Type Plasminogen Activator (*PLAT*), and Protease Nexin 1 (*SERPINE2*)¹⁰.

In the original publication on CP-GEP¹⁰, performance was stratified by tumor (T) stage because currently, T staging constitutes the basis of SLNB decision making. The predictive value of CP-GEP over established staging parameters was expressed by the SLNB reduction rate, a metric introduced by Mocellin and colleagues³³ to quantify the fraction of patients who can be deselected for SLNB by a test. The SLNB reduction rate of CP-GEP varied by T stage, was highest for T1b melanoma, and then decreased as tumor thickness increased¹⁰. Of note, the SLNB reduction rate remained positive even for T3b melanoma, indicating that CP-GEP outperformed CP variables across a broad range of tumor thickness. Bartlett and colleagues⁴⁶ from the MSKCC acknowledged in a critical review that CP-GEP achieved a 14% improvement in SLNB reduction rate over the best CP models. The overall SLNB reduction rate over current clinical practice, which is based on T staging, was ~60%¹⁰.

While achieving a high SLNB reduction rate is desirable, test results need to be oncologically safe. In other words, the number of false-negative test results for a rule-out test like CP-GEP needs to be <5% and the NPV > 95%. These are the metrics that apply to current patient selection. SLNB should be considered and discussed with patients if Breslow thickness is ≥ 0.8 mm (stage T1b) because T1b melanoma metastasizes to SLNB at a rate of $\geq 5\%$. SLNB is recommended for T2a melanoma because T2a melanoma metastasizes to SLNB at rate of $\geq 10\%$. T1a melanoma, in contrast, metastasizes at a rate below 5% and, therefore, does not require SLNB¹. At a false-negative rate of <5% for T1a melanoma, we likely miss a non-trivial number of node positive T1a patients because the number of patients with T1a melanoma is very large. But as part of the trade-off, we also avoid a very large number of unnecessary, non-therapeutic SLNB procedures. The idea of a test like CP-GEP is to extract molecular information from slightly thicker melanomas, such as T1b and T2 melanomas, and use molecular information to push the pretest probability of a positive SLNB to below 5% (NPV > 95%). Low risk tested patients can then forgo SLNB based on established patient selection criteria, which allows us to dramatically reduce the number of unnecessary SLNB. However, because we stop doing SLNB on all patients with T1b and T2 melanoma, we also miss a few patients with positive SLNB. The trade-off is the same as for T1a melanoma.

Some have argued that ruling-out patients for SLNB by CP-GEP is problematic because patients with false-negative test results are precluded from receiving adjuvant systemic therapy⁴⁶. However, the likelihood of receiving a false negative test result with CP-GEP is no higher than for T1a melanoma, which we also do not consider for SLNB. Further, CP-GEP testing might aid in the interpretation of SLNB with minimal tumor burden. More than 20% of patients with positive SLNB have single positive nodes with minimal numbers of melanocytes (individual tumor cells or cell clusters <0.1 mm)^{10,47}, which are often only detectable by highly sensitive methods such as immunohistochemistry. The prognostic significance of minimal SLN tumor burden is unclear and optimal treatment uncertain. Preliminary evidence from an international multiinstitutional cohort of 1,684 melanoma patients suggests that CP-GEP might help stratify patients with minimal SLN tumor burden according to recurrence risk and guide clinical decision-making beyond nodal assessment⁴⁸. The prognostic value of CP-GEP has previously been established by Eggermont et al. who have shown that CP-GEP very effectively stratifies SLNB-negative patients based on their relapse risk^{45,49,50}.

During the COVID-19 pandemic, CP-GEP has helped prioritize patients for surgery, thereby optimizing healthcare resources^{20,51}. CP-GEP is the only test with a GEP component that has been shown to add predictive value over complex CP models such as the MSKCC¹⁰ and MIA nomograms⁴⁴. A multicenter registry study is ongoing in the United States to prospectively validate CP-GEP⁵². A number of retrospective validation studies have also been published^{47,53–55}.

MOLECULAR TESTING: WHY BOTHER?

Some authors have argued that CP-GEP adds cost to patient care without significantly outperforming CP-based models like the MIA nomogram⁵⁶. Here, we argue for molecular testing and discuss the limitations of models that solely rely on CP-based nomograms.

Issue #1: There is variability in the reporting of CP variables used in nomograms

Patient age, Breslow thickness, tumor ulceration, and TNM pathological stages are consistently reported among trained pathologists from different centers and geographical areas. There is less agreement on other histopathological features, including Clark level, regression, TILs, MR, and LVI, which are often used in CP-based nomograms^{27,28,57,58}, or experimental variables such as

calculated tumor area, which have yet to be validated⁵⁹. As was pointed out by Monshizadeh and colleagues²⁷, the majority of melanomas are diagnosed in non-specialized non-academic centers, and variability in reporting is a function of training and experience. In the United States, there are disparities in the reporting of MR across institutions that exist within traditionally disadvantaged populations⁶⁰. Of note, Hispanics and non-Whites are also less likely to undergo excisional biopsies⁶¹. Because of the variability in histopathology read-out, some dermatopathologists have argued for the routine use of GEP-based testing, which they consider methodologically precise⁶².

Issue #2: CP-based nomograms produce inconsistent results and have not been extensively validated

Three CP-based SLNB prediction models from specialized academic centers are accessible online: the MSKCC^{31,63}, MIA^{32,64} and Life Math (Harvard) nomograms⁶⁵. These models use different sets of predictors. For example, the newer MIA model considers LVI, whereas the older MSKCC and Life Math models do not. Not all patients fit the preset categorizations of nominal variables like histologic type in some of the nomograms. Conversely, for MR, models require discrete values when pathologists often report ordinal ranges. In our experience, MIA nomogram risk scores could not be calculated for 10 to 20% of patients, mostly because of incompatible histologic types^{44,54}. Moreover, there are disparities in predicted outcomes between the nomograms that directly impact patient care⁶⁶. These are likely due to variable choice, historical differences between model development cohorts, and local idiosyncrasies of pathology interpretation.

Expectations expressed by melanoma key opinion leaders for the development and validation of GEP-based tools⁴⁰ greatly exceed what has been the norm for CP-based nomograms. 31-GEP leads the way in the number of validation studies and publications, however, clinical and statistical experts have criticized data quality, study design and statistical approach^{67–69}. A multicenter clinical trial is underway in Germany to study 11-GEP for postoperative adjuvant therapy in stage II patients⁴³. CP-GEP has been validated retrospectively in multiple independent cohorts both in the United States and Europe^{47,53–55}. A multicenter prospective registry study for CP-GEP is recruiting in the United States⁵².

Issue #3: Molecular testing has the potential to be cost saving

For patients with a pretest probability of SLNB positivity that is greater than 5–10% and who do not otherwise qualify for adjuvant pembrolizumab⁸, SLNB continues to be critical for therapeutic decision making¹. However, if patient selection is solely based on Breslow thickness and tumor ulceration, as is currently the case¹, approximately 85% of SLNB procedures will be negative and non-therapeutic¹⁰. To better quantify the pretest probability of SLNB metastasis and reduce the number of SLNB-eligible patients without sacrificing oncologic safety, molecular testing will likely have a role to play. Whether molecular testing is cost saving depends on the cost of the test, the cost of SLNB, and the SLNB reduction rate that a test can achieve while maintaining oncologic safety at an NPV of ~95%. It follows that the great majority of T1a patients should not be tested as their risk of nodal metastasis is <5% based on Breslow thickness alone. Exceptions may include patients with transected biopsies, elevated MR, or other worrisome histopathologic features recognized by an experienced dermatopathologist. T1b and T2 melanomas, in contrast, are well-suited for a test like CP-GEP because the SLNB reduction rate achieved at an oncologically safe NPV of ~95% is high (up to 80%)¹⁰. At an average cost of SLNB in the United States of ~\$19,000⁷⁰, CP-GEP testing would be cost-saving even at a several thousand dollar price point. Studies on this important topic are forthcoming. Of note, molecular testing not only reduces the rate of SLNB but also improves health-related quality

of life as CP-GEP low-risk patients can forgo a complex surgical procedure²⁰.

FUTURE DIRECTIONS

Melanoma risk assessment is changing with the recent advent of molecular tests like CP-GEP, which hold the promise of improving patient selection for SLNB and other interventions¹⁰. We interpret these tests as an important step forward, as they hold the promise of improving patient outcomes while reducing healthcare costs. Molecular tests like CP-GEP might help us avoid negative, non-therapeutic SLNB so that healthcare resources can be reallocated to more deserving interventions^{20,51}. CP-based nomograms, which have been promoted as alternatives to molecular testing, tend to be unreliable as they rely on complex CP features such as TILs, MR, and LVI. These features are inconsistently reported among pathologists, which has been attributed to variability in training, experience, the local quality of care, and idiosyncrasies of histopathology interpretation^{27,61,66}. It is therefore unsurprising that CP nomograms perform well in development cohorts from world-renowned subspecialized centers³² but validate poorly in alternate settings⁵⁴. Molecular tests on the other hand are methodologically precise across continents and patient populations. Such tests can be combined with clear-cut CP predictors like Breslow thickness and patient age^{10,71}. As we await the transition to a molecular-driven world, the careful evaluation of melanoma histopathology and sentinel nodes will remain important cornerstones of melanoma staging and therapy.

REFERENCES

- Swetter SM, Thompson JA, Albertini MR, Barker CA, Baumgartner J, Boland G, et al. NCCN Guidelines® Insights: Melanoma: Cutaneous, Version 2.2021: Featured Updates to the NCCN Guidelines. *J. Natl. Compr. Canc. Netw.* **19**, 364–376 (2021).
- Gershenwald JE, Scolyer RA, Hess KR, Sondak VK, Long GV, Ross MI, et al. Melanoma staging: evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA Cancer J. Clin.* **67**, 472–492 (2017).
- Ross MI, Reintgen D, Balch CM. Selective lymphadenectomy: emerging role for lymphatic mapping and sentinel node biopsy in the management of early stage melanoma. *Semin. Surg. Oncol.* **9**, 219–223 (1993).
- Galmarini CM. Why we do what we do. A brief analysis of cancer therapies. *EXCLI J.* **19**, 1401–1413 (2020).
- Faries MB, Thompson JF, Cochran AJ, Andtbacka RH, Mozzillo N, Zager JS, et al. Completion Dissection or Observation for Sentinel-Node Metastasis in Melanoma. *N. Engl. J. Med.* **376**, 2211–2222 (2017).
- Leiter U, Stadler R, Mauch C, Hohenberger W, Brockmeyer N, Berking C, et al. Complete lymph node dissection versus no dissection in patients with sentinel lymph node biopsy positive melanoma (DeCOG-SLT): a multicentre, randomised, phase 3 trial. *Lancet Oncol.* **17**, 757–767 (2016).
- Rentiroia-Pacheco B, Tijen-Fooh FJ, Quattrocchi E, Kobic A, Wever R, Bellomo D, et al. Clinicopathologic models predicting non-sentinel lymph node metastasis in cutaneous melanoma patients: Are they useful for patients with a single positive sentinel node? *J. Surg. Oncol.* **125**, 516–524 (2022).
- Luke JJ, Rutkowski P, Queirolo P, Del Vecchio M, Mackiewicz J, Chiarion-Sileni V, de la Cruz Merino L, Khattak MA, Schadendorf D, Long GV, Ascierto PA, Mandalà M, De Galitiis F, Haydon A, Dummer R, Grob JJ, Robert C, Carlino MS, Mohr P, Poklepovic A, Sondak VK, Scolyer RA, Kirkwood JM, Chen K, Diede SJ, Ahsan S, Ibrahim N, Eggermont AMM. KEYNOTE-716 Investigators. Pembrolizumab versus placebo as adjuvant therapy in completely resected stage IIB or IIC melanoma (KEYNOTE-716): a randomised, double-blind, phase 3 trial. *Lancet.* **399**, 1718–1729 (2022).
- Morton DL, Thompson JF, Cochran AJ, Mozzillo N, Nieweg OE, Roses DF, et al. Final Trial Report of Sentinel-Node Biopsy versus Nodal Observation in Melanoma. *N. Engl. J. Med.* **370**, 599–609 (2014).
- Bellomo D, Arias-Mejias SM, Ramana C, Heim JB, Quattrocchi E, Somnidi-Damodaran S, et al. Model Combining Tumor Molecular and Clinicopathologic Risk Factors Predicts Sentinel Lymph Node Metastasis in Primary Cutaneous Melanoma. *JCO Precis Oncol.* **4**, 319–334 (2020).
- Moody JA, Ali RF, Carbone AC, Singh S, Hardwicke JT. Complications of sentinel lymph node biopsy for melanoma - A systematic review of the literature. *Eur. J. Surg. Oncol.* **43**, 270–277 (2017).

12. Hieken TJ, Sadurní MB, Quattrocchi E, Kobic A, Somnidi-Damodaran S, Dwarkasing JT, et al. Using the Merlin assay for reducing sentinel lymph node biopsy complications in melanoma: a retrospective cohort study. *Int. J. Dermatol.* <https://doi.org/10.1111/ijd.16056> (2022).
13. Biver-Dalle C, Puzenat E, Puyraveau M, Delroeu D, Boulahdour H, Sheppard F, et al. Sentinel lymph node biopsy in melanoma: Our 8-year clinical experience in a single French institute (2002–2009). *BMC Dermatol.* **12**, 21 (2012).
14. Lee RJ, Gibbs JF, Proulx GM, Kollmorgen DR, Jia C, Kraybill WG. Nodal basin recurrence following lymph node dissection for melanoma: implications for adjuvant radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* **46**, 467–474 (2000).
15. Nieweg OE, Estourgie SH. What is a sentinel node and what is a false-negative sentinel node? *Ann Surg Oncol.* **11**, 1695–735 (2004).
16. Brennick JB, Yan S. False-positive cells in sentinel lymph nodes. *Semin. Diagn. Pathol.* **25**, 116–119 (2008).
17. Yan S, Brennick JB. False-positive rate of the immunoperoxidase stains for MART1/MelanA in lymph nodes. *Am. J. Surg. Pathol.* **28**, 596–600 (2004).
18. Hayek SA, Munoz A, Dove JT, Hunsinger M, Arora T, Wild J, et al. Hospital-Based Study of Compliance with NCCN Guidelines and Predictive Factors of Sentinel Lymph Node Biopsy in the Setting of Thin Melanoma Using the National Cancer Database. *Am. Surg.* **84**, 672–679 (2018).
19. Glas AM, Floore A, Delahaye LJM, Witteveen AT, Pover RCF, Bakx N, et al. Converting a breast cancer microarray signature into a high-throughput diagnostic test. *BMC Genomics* **7**, 278 (2006).
20. Meves A, Eggermont AMM. Deselecting Melanoma Patients for Sentinel Lymph Node Biopsy During COVID-19: Clinical Utility of Tumor Molecular Profiling. *Mayo Clin Proc Innov Qual Outcomes* **4**, 586–587 (2020).
21. Balch CM. Measuring melanomas—a tribute to Alexander Breslow. *J. Am. Acad. Dermatol.* **5**, 96–97 (1981).
22. Pickett H. Shave and punch biopsy for skin lesions. *Am. Fam. Physician* **84**, 995–1002 (2011).
23. de Menezes SL, Kelly JW, Mar V. The increasing use of shave biopsy for diagnosing invasive melanoma in Australia. *Med. J. Aust.* **212**, 238–238.e1 (2020).
24. Balch CM, Thompson JF, Gershenwald JE, Soong S-J, Ding S, McMasters KM, et al. Age as a predictor of sentinel node metastasis among patients with localized melanoma: an inverse correlation of melanoma mortality and incidence of sentinel node metastasis among young and old patients. *Ann. Surg. Oncol.* **21**, 1075–1081 (2014).
25. Balch CM, Soong S-J, Gershenwald JE, Thompson JF, Coit DG, Atkins MB, et al. Age as a prognostic factor in patients with localized melanoma and regional metastases. *Ann. Surg. Oncol.* **20**, 3961–3968 (2013).
26. Hawryluk EB, Pappo AS, Marghoob AA, Bahrami A, Tsao H. Melanoma in children. *UpToDate*. Hensin Tsao (Ed). *UpToDate*, Wolters Kluwer (2021). Available from: <https://www.uptodate.com/contents/101324>.
27. Monshizadeh L, Hanikeri M, Beer TW, Heenan PJ. A critical review of melanoma pathology reports for patients referred to the Western Australian Melanoma Advisory Service. *Pathology* **44**, 441–447 (2012).
28. Patrawala S, Maley A, Greskovich C, Stuart L, Parker D, Swerlick R, et al. Discordance of histopathologic parameters in cutaneous melanoma: Clinical implications. *J. Am. Acad. Dermatol.* **74**, 75–80 (2016).
29. Elmore JG, Elder DE, Barnhill RL, Knezevich SR, Longton GM, Titus LJ, et al. Concordance and Reproducibility of Melanoma Staging According to the 7th vs 8th Edition of the AJCC Cancer Staging Manual. *JAMA Netw Open* **1**, (2018).
30. Cheraghlou S, Ugwu N, Girardi M. Sentinel Lymph Node Biopsy Positivity in Patients With Acral Lentiginous and Other Subtypes of Cutaneous Melanoma. *JAMA Dermatol.* (2021).
31. Risk of sentinel lymph node metastasis [Internet]. [cited 10 December 2021]. Available from: https://www.mskcc.org/nomograms/melanoma/sentinel_lymph_node_metastasis.
32. Lo SN, Ma J, Scolyer RA, Haydu LE, Stretch JR, Saw RPM, et al. Improved Risk Prediction Calculator for Sentinel Node Positivity in Patients With Melanoma: The Melanoma Institute Australia Nomogram. *J. Clin. Oncol.* **38**, 2719–2727 (2020).
33. Mocellin S, Thompson JF, Pasquali S, Montesco MC, Pilati P, Nitti D, et al. Sentinel Node Status Prediction by Four Statistical Models: Results From a Large Bi-Institutional Series (n = 1132). *Ann. Surg.* **250**, 964 (2009).
34. Meves A, Nikolova E, Heim JB, Squirewell EJ, Cappel MA, Pittelkow MR, et al. Tumor Cell Adhesion As a Risk Factor for Sentinel Lymph Node Metastasis in Primary Cutaneous Melanoma. *J. Clin. Oncol.* **33**, 2509–2515 (2015).
35. Zanivan S, Meves A, Behrendt K, Schoof EM, Neilson LJ, Cox J, et al. In vivo SILAC-based proteomics reveals phosphoproteome changes during mouse skin carcinogenesis. *Cell Rep.* **3**, 552–566 (2013).
36. Kashani-Sabet M, Rangel J, Torabian S, Nosrati M, Simko J, Jablons DM, et al. A multi-marker assay to distinguish malignant melanomas from benign nevi. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 6268–6272 (2009).
37. Brunner G, Reitz M, Heinecke A, Lippold A, Berking C, Suter L, et al. A nine-gene signature predicting clinical outcome in cutaneous melanoma. *J. Cancer Res. Clin. Oncol.* **139**, 249–258 (2013).
38. Gerami P, Cook RW, Wilkinson J, Russell MC, Dhillon N, Amaria RN, et al. Development of a prognostic genetic signature to predict the metastatic risk associated with cutaneous melanoma. *Clin. Cancer Res.* **21**, 175–183 (2015).
39. Amaral TMS, Hoffmann M-C, Sinnberg T, Niessner H, Sülberg H, Eigentler TK, et al. Clinical validation of a prognostic 11-gene expression profiling score in prospectively collected FFPE tissue of patients with AJCC v8 stage II cutaneous melanoma. *Eur. J. Cancer* **125**, 38–45 (2020).
40. Grossman D, Okwundu N, Bartlett EK, Marchetti MA, Othous M, Coit DG, et al. Prognostic Gene Expression Profiling in Cutaneous Melanoma: Identifying the Knowledge Gaps and Assessing the Clinical Benefit. *JAMA Dermatol.* **156**, 1004–1011 (2020).
41. Brunner G, Heinecke A, Falk TM, Ertaş B, Blödorn-Schlicht N, Schulze H-J, et al. A Prognostic Gene Signature Expressed in Primary Cutaneous Melanoma: Synergism With Conventional Staging. *JNCI Cancer Spectr* **2**, ky032 (2018).
42. MolDX: Melanoma Risk Stratification Molecular Testing [Internet]. Centers for Medicare & Medicaid Services. Available from: <https://www.cms.gov/medicare-coverage-database/search.aspx>.
43. Adjuvant Nivolumab Treatment in Stage II (IIA, IIB, IIC) High-risk Melanoma - Full Text View - ClinicalTrials.gov [Internet]. [cited 12 December 2021]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04309409>.
44. Bellomo D, Bridges AG, Hieken TJ, Meves A. Reply to E. K. Bartlett et al and A. H. R. Valey et al. *JCO Precis. Oncol.* 992–994 (2020).
45. Eggermont AMM, Bellomo D, Dwarkasing J, Meerstein-Kessel L, Meves A. The letter responds to comment on Identification of stage I/IIA melanoma patients at high risk of disease relapse using a clinicopathologic and gene expression model. *Eur. J. Cancer* (2021).
46. Bartlett EK, Marchetti MA, Coit DG. Gene Expression Profile-Based Risk Modeling to Select Patients With Melanoma Who Can Avoid Sentinel Lymph Node Biopsy: Are We There Yet? *JCO Precis. Oncol.* 988–989 (2020).
47. Yousaf A, Tjien-Fooh FJ, Rentroia-Pacheco B, Quattrocchi E, Kobic A, Tempel D, et al. Validation of CP-GEP (Merlin Assay) for predicting sentinel lymph node metastasis in primary cutaneous melanoma patients: A U.S. cohort study. *Int. J. Dermatol.* **60**, 851–856 (2021).
48. Tjien-Fooh F, Meves A, Wever R, Hollestein L, Mulder E, Johansson I, et al. Cutaneous melanoma patients with minimal SN tumor burden: CP-GEP (Merlin Assay) may guide decision-making beyond nodal assessment. *J. Am. Acad. Dermatol.* in press.
49. Eggermont AMM, Bellomo D, Arias-Mejias SM, Quattrocchi E, Somnidi-Damodaran S, Bridges AG, et al. Identification of stage I/IIA melanoma patients at high risk for disease relapse using a clinicopathologic and gene expression model. *Eur. J. Cancer* **140**, 11–18 (2020).
50. Wever R, Tjien-Fooh F, Bellomo D, Quattrocchi E, Damodaran SS, Van Vliet M, et al. Identification of stage IIA melanoma patients at high risk for disease relapse using a clinicopathologic and gene expression model. *J. Clin. Oncol.* **38**, e22088–e22088 (2020).
51. Mulder EEAP, Tempel D, Mooyart AL, Verhoef C, Dwarkasing JT, Grunhagen DJ. Using the clinicopathologic and gene expression (CP-GEP) model to predict sentinel node status in patients with primary melanoma: A prospective cohort study during the COVID-19 pandemic [Internet]. *falconprogram.com*. [cited 22 June 2021]. Available from: https://falconprogram.com/files/poster_P108_EADO_Conference_2021.pdf.
52. Melanoma Research Lymph Node Prediction Implementation National_001 - Full Text View - ClinicalTrials.gov [Internet]. [cited 12 December 2021]. Available from: <https://clinicaltrials.gov/ct2/show/NCT04759781>.
53. Mulder EEAP, Dwarkasing JT, Tempel D, van der Spek A, Bosman L, Verver D, et al. Validation of a clinicopathologic and gene expression profile model for sentinel lymph node metastasis in primary cutaneous melanoma. *Br. J. Dermatol.* (2020).
54. Johansson I, Tempel D, Dwarkasing JT, Rentroia-Pacheco B, Mattsson J, Ny L, et al. Validation of a clinicopathologic and gene expression profile model to identify patients with cutaneous melanoma where sentinel lymph node biopsy is unnecessary. *Eur. J. Surg. Oncol.* (2021).
55. Arias-Mejias SM, Quattrocchi E, Tempel D, Luna-Vargas M, Chen J, Murphree DH, et al. Primary cutaneous melanoma risk stratification using a clinicopathologic and gene expression model: a pilot study. *Int. J. Dermatol.* **59**, e431–e433 (2020).
56. Valey AHR, Lo SN, Scolyer RA, Thompson JF. Predicting Sentinel Node Status in Patients With Melanoma: Does Gene Expression Profiling Improve Accuracy? *JCO Precision Oncology*, 990–991 (2020).
57. Barnhill RL, Busam KJ, From L, Bagot M, Lugassy C, Berwick M. Inter-observer concordance for the recognition of angiotropism in human melanoma. *Pigment Cell Melanoma Res.* **24**, 582–583 (2011).
58. Saldanha G, Ali R, Bakshi A, Basiouni A, Bishop R, Colloby P, et al. Global and mitosis-specific interobserver variation in mitotic count scoring and implications for malignant melanoma staging. *Histopathology* **76**, 803–813 (2020).

59. Saldanha G, Yarrow J, Elsheikh S, O'Riordan M, Uraiby H, Bamford M. Development and initial validation of calculated tumor area as a prognostic tool in cutaneous malignant melanoma. *JAMA Dermatol.* **155**, 890–898 (2019).
60. Lorimer PD, Benham EC, Walsh K, Han Y, Forster MR, Sarantou T, et al. Reporting of mitotic rate in cutaneous melanoma: A study using the national cancer data base. *J. Surg. Oncol.* **115**, 281–286 (2017).
61. Restrepo DJ, Huayllani MT, Boczar D, Sisti A, Gabriel E, Lemini R, et al. Biopsy Type Disparities in Patients With Melanoma: Who Receives the Standard of Care? *Anticancer Res.* **39**, 6359–6363 (2019).
62. Glazer A, Cockerell C. Histopathologic discordance in melanoma can have substantial impacts on patient care. *J of Skin* **3**, 85–89 (2019).
63. Wong SL, Kattan MW, McMasters KM, Coit DG. A nomogram that predicts the presence of sentinel node metastasis in melanoma with better discrimination than the American Joint Committee on Cancer staging system. *Ann. Surg. Oncol.* **12**, 282–288 (2005).
64. Melanoma Risk Assessment Tool [Internet]. [cited 13 December 2021]. Available from: <https://www.melanomarisk.org.au/SNLForm>.
65. Melanoma nodal status calculator [Internet]. [cited 13 December 2021]. Available from: <http://lifemath.net/cancer/melanoma/nodal/index.php>.
66. Butt ST, Chin KY, Proby CM, Affleck AG. Prediction of sentinel node status using melanoma e-prognostic tools. *Clin. Exp. Dermatol.* **46**, 743–744 (2021).
67. Kovarik CL, Chu EY, Adamson AS. Gene Expression Profile Testing for Thin Melanoma. *JAMA Dermatology.* **156**, 837 (2020).
68. Marchetti MA, Bartlett EK, Dusza SW, Bichakjian CK. Use of a prognostic gene expression profile test for T1 cutaneous melanoma: Will it help or harm patients? *J. Am. Acad. Dermatol.* **80**, e161–e162 (2019).
69. Marchetti MA, Dusza SW, Bartlett EK. Problematic methodology in a systematic review and meta-analysis of DecisionDx-Melanoma. *J. Am. Acad. Dermatol.* **83**, e357–e358 (2020).
70. Marson J, Quick A, Martin B, Litchman G, Rigel D. The clinical and financial impact of the 31-gene expression profile testing on sentinel lymph node biopsy patients selection in patients with T1b cutaneous melanoma. Poster presentation. 19th Annual South Beach Symposium, Miami Beach (2021).
71. Whitman ED, Koshenkov VP, Gastman BR, Lewis D, Hsueh EC, Pak H, et al. Integrating 31-Gene Expression Profiling With Clinicopathologic Features to Optimize Cutaneous Melanoma Sentinel Lymph Node Metastasis Prediction. *JCO Precis. Oncol.* **5**, <https://doi.org/10.1200/PO.21.00162> (2021).

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M.B.S. and A.M. reviewed the literature and wrote the manuscript. Both authors read and approved the final manuscript.

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COMPETING INTERESTS

Mayo Clinic and AM have a financial interest in the CP-GEP (Merlin) Assay. AM received research funding from SkylineDx. Ms. Sadurní has no conflicts of interest to declare.

ADDITIONAL INFORMATION

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